

Curcumin and Cancer: Recent Developments

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ABSTRACT:

Cancer is a malignant disease with high mortality rates affecting millions. Although chemotherapeutic agents are employed largely in cancer management, they often result in toxicity and side effects and may lead to resistance. These drawbacks of the conventional chemotherapeutic agents have led to the urgent need for development of safer, biocompatible, nontoxic compounds from natural sources and their application in cancer management. Products from natural sources are being exploited in cancer research worldwide due to their less toxicity. Of the several natural products tested, curcumin, well known for its chemopreventive, cytoprotective and immune suppressive properties holds great promise for cancer research. Curcumin has been reported to affect different signaling pathways either in a direct or indirect manner in a wide range of cancers. Despite the therapeutic effectiveness of curcumin, its application is largely restricted due to its poor absorption, lipophilic nature and low bioavailability. Thus newer and effective formulations for curcumin in cancer treatment are being continuously exploited. In this review we highlighted (i) recent developments of application of natural products in cancer research (ii) role of curcumin in different cancers (iii) curcumin formulations and their application in cancer research. The future scope of this review lies in the effective employment of curcumin and its formulations, in the eradication of cancer.

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INTRODUCTION

Despite ongoing progress of the chemotherapeutic agents in cancer management, it still remains to be a major killer. The main challenge in cancer therapeutics revolves around specific killing of cancer cells within a milieu of the normal ones, overcome resistance to chemotherapeutic drugs called as Multi Drug Resistance (MDRs) and prevention of metastasis, targeting clinical relapse and augmentation of chemotoxicity. The conventional anticancer agents employed in cancer management are mostly toxic that lead to life threatening complications of neurotoxicity and cognitive changes (Dutta *et al.*, 2011; Fardell *et al.*, 2011). Therefore the need for safer, nontoxic products remains in targeting cancer. Different types of natural products are being continuously exploited in cancer research (Ulbricht *et al.*, 2010; Dai *et al.*, 2010) due to the tested advantage of being less toxic than synthesized chemical compounds.

Curcumin, a major curcuminoid derivative, is a main component of turmeric obtained from the roots of *Curcuma longa* and forms an important component of the South Asian cuisine. (Fig-1). It is chemically formulated as diferuloylmethane ($C_{21}H_{20}O_6$), a polyphenol in its properties, curcumin has been reported to affect the biological system immensely by its antiinflammatory, antitumorigenic, antioxidant, antiseptic, anti-toxic, cancer chemopreventive, chemo sensitization, radio sensitization epigenetic change inducer, and potentially chemotherapeutic properties (Aggarwal *et al.*, 2007, Basnet *et al.*, 2011) and thus finds wide application in health research.

Several classes of bio-molecules are selectively regulated by curcumin thereby affecting diverse signaling pathways (Arora *et al.*, 1973; Srimal *et al.*, 1973; Jobin *et al.*, 1999; Sharma *et al.*, 2005 and Shanmugam *et al.*, 2011) in the biological system.

The hall mark property of targeted cell killing by curcumin (Shanmugam *et al.*, 2011; Syng-Ai *et al.*,

2004), through different signaling pathways portray it as a promising chemotherapeutic molecule. The exact molecular mechanism of curcumin in different cancer cells is being determined. However some of the target molecules that curcumin effect in different cancers include nuclear factor- κ B, activator protein-1, nitric oxide synthase, receptor tyrosine kinases, cell cycle regulators like cyclins, matrix metalloproteinases, proapoptotic markers and inhibits angiogenesis (Syng-Ai *et al.*, 2004; Aggarwal *et al.*, 2007; Bierhaus *et al.*, 1997; Brouet *et al.*, 1995; Hahm *et al.*, 2004; Korutla *et al.*, 1994; Singh *et al.*, 1995). Curcumin is currently in clinical trials for treatment of various cancers, including multiple myeloma, pancreatic cancer, and colon cancer (Aggarwal *et al.*, 2003).

In this review we focus our attention to (a) mortality rates in different cancer (b) recent references to natural products applied in cancer research (c) how curcumin inhibits cancer through different pathways (d) recent developments in curcumin formulations and applications in cancer research.



Fig 1A: Plant



Fig 1C: Rhizome

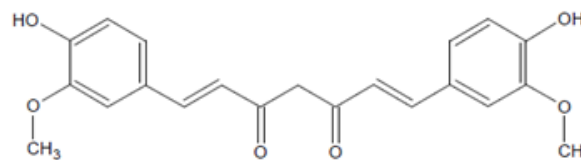


Fig 1B: Chemical Structure of curcumin

Fig 1: Curcumin: 1A. Plant, 1B. Chemical Structure of curcumin, 1C. Rhizome



Mortality rates in different types of cancer

Cancer remains to be a leading cause of death worldwide accounting for 7.6 million deaths (around 13% of all deaths) in 2008, as determined by statistics on Cancer Incidence and Mortality Worldwide by GLOBOCAN, Cancer Fact Sheet conducted by WHO (<http://www.globocon.iarc.fr/>) (Fig-2). The statistics summarized the mortality rate in cancer and was represented by an Age-standardized rate (W) including the number of new cases or deaths per 100,000 persons per year. In India the most frequent cancers reported were cervix and uteri, breast, oral and lips, lungs and oesophagus while lung, breast, colorectal and prostate were the five most frequent occurring ones in the world.

Hence the need for suitable molecules to fight the disease is urgent.

Recent references of Natural products applied in cancer research

A whole range of natural products obtained from bacterial, algal, fungal and higher plant sources (Kinghorn et al., 2011) are being exploited for obtaining active molecules to combat cancer (Table-1).

Curcumin: Molecular Pathways and Inhibition of Cancer

Cancer is a disease with major manifestations like uncontrolled cell proliferation and metastasis. Research in the field of cancer chemotherapy broadly revolves around the control by the following mechanisms

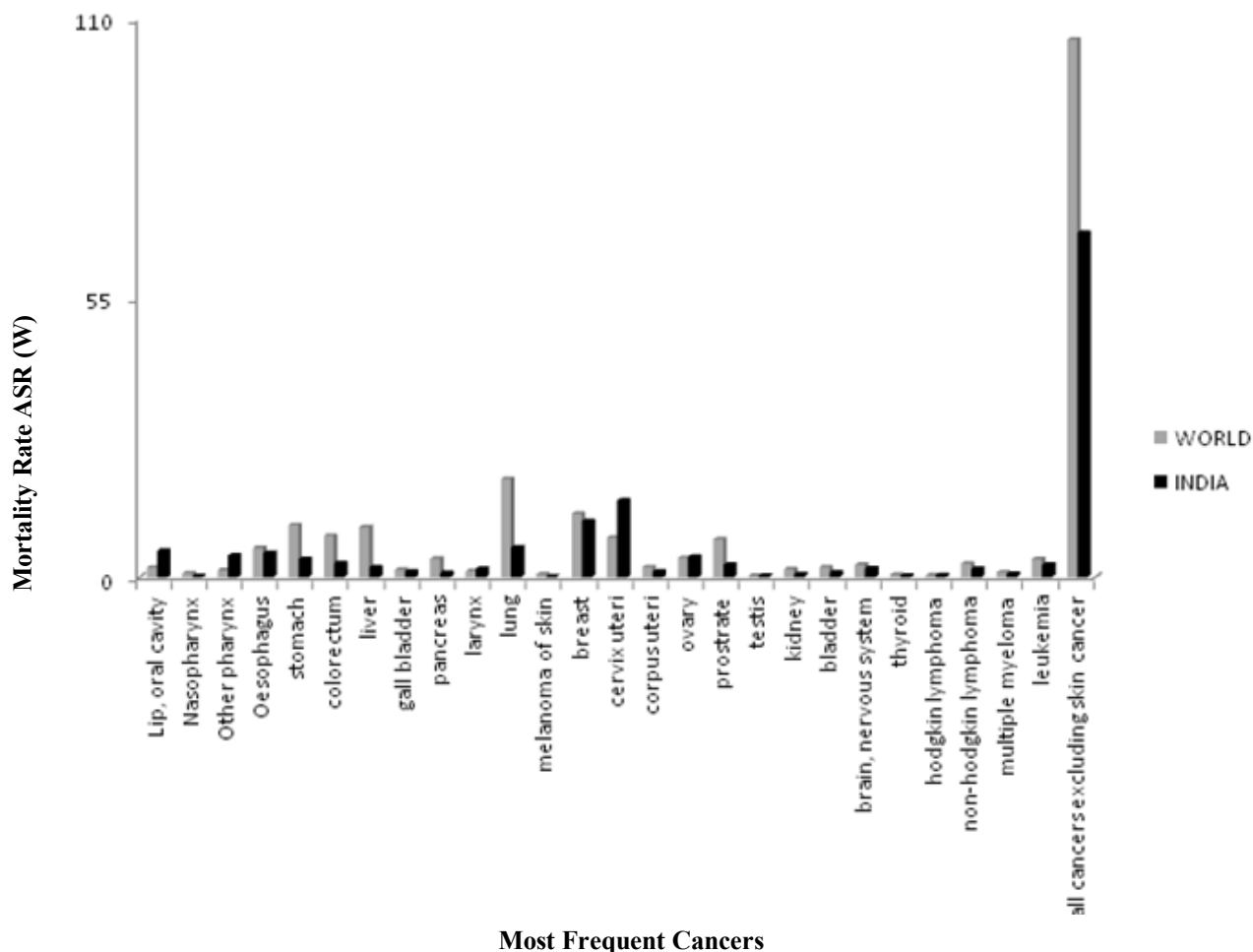


Fig 2: Cancer Statistics: World and India: The statistics is based on Mortality in most frequent cancers. The data is based on Age-standardized rate (W): A rate is the number of new cases or deaths per 100, 000 persons per year. An age-standardized rate is the rate that a population would have if it had a standard age structure. (Globocon, WHO 2008, Cancer Facts Sheet).

(i) Inhibition of cell proliferation and metastasis (ii) Induced apoptosis (iii) Overcoming MDRs, (iv) Tumour suppression. The effects of curcumin, in inducing the above roles both by direct and indirect effect on molecules have been extensively studied in different cancer and cell lines (**Fig-3**). Curcumin has been reported to play a dominant role in the prevention of different types of cancer by promoting apoptosis or by inhibiting cell proliferation.

In pituitary cancer, curcumin has been reported to inhibit cell proliferation (Schaaf *et al.*, 2009; Schaaf *et al.*, 2010) by decreased cyclin D3 expression thus affecting cell cycle G₁ to S transition and suppressed growth hormone (GH) levels like chymothypsin prolactins and enhanced the growth-inhibitory effect of low concentrations of bromocriptine. It is also known to induce apoptosis by decreased phosphorylation of

retinoblastoma protein (Rb), and block clonogenicity of tumor cells (Miller *et al.*, 2008) thus enabling cancer prevention.

In colon carcinoma, curcumin inhibits tumor growth by increased apoptosis through the expression of cyclooxygenase-2 (COX-2) and inhibition of proteasomal chymothypsin-like activity thereby leading to accumulation of ubiquitinated proteins and proteasome target proteins IκB-α, p27, and p21/Bax leading to apoptosis (Milacic *et al.*, 2008), activation of Reactive Oxygen Species (ROS) (Lee YJ *et al.*, 2011) and, suppression of mitochondrial NADP(+)-dependent isocitrate dehydrogenase activity (Jung KH *et al.*, 2011). Curcumin also inhibits tumor cell proliferation by promoting cell cycle arrest in the G₁ phase manifested by decreased levels of PCNA, Cyclin D1, C-Myc, and Bcl-2, Nuclear Factor kappa B, NF-κB-regulated

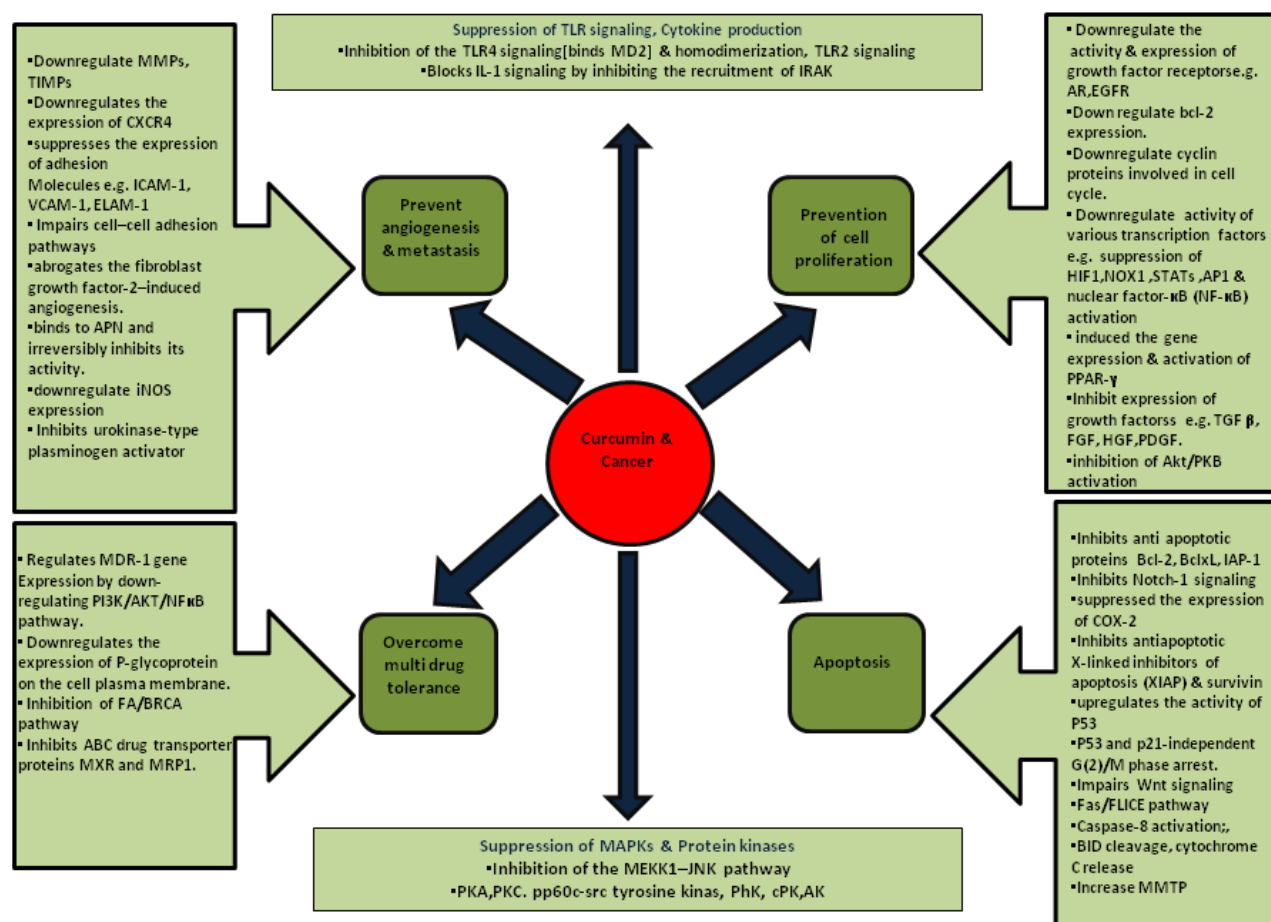


Fig 3: Curcumin and Cancer: Biomolecules and Pathways



Table 1: Natural Products and Cancer Research: Recent Developments

Major source of Natural Product	Examples and References
Bacteria	Bacterial products (Wang C <i>et al.</i> , 2011), Largazole, from the marine cyanobacterium <i>Symploca</i> species (Li S <i>et al.</i> , 2011), Marine cyanobacterium <i>Lyngbya majuscula</i> (Tripathi <i>et al.</i> , 2011).
Algae	Red sea weeds (Ahmed <i>et al.</i> , 2011), Alga (Shalaby <i>et al.</i> , 2011)
Fungus	Endophytic fungal strain(Wang XN <i>et al.</i> , 2011), Halophilic actinomycete <i>Actinopolyspora erythraea</i> YIM 90600(Zhao <i>et al.</i> , 2011), Mushroom(Jiang <i>et al.</i> , 2010).
Herbs	Chinese herbs (Eichhorn <i>et al.</i> , 2011), Thunder god vine or <i>Tripterygium wilfordii</i> Hook. F. (Liu Z <i>et al.</i> , 2011), Herbal flavenoids(Liu PX <i>et al.</i> , 2011), lipid-soluble ginseng extract (LSGE)(Kang <i>et al.</i> , 2011), <i>Phyllanthus urinaria</i> (Huang ST <i>et al.</i> , 2010).
Higher plants	Curcumin(Aggarwal <i>et al.</i> , 2007; Basnet <i>et al.</i> , 2011), Rhizome of <i>Cnidium officinale</i> (Bae <i>et al.</i> , 2011), <i>Hypoestes forskoolii</i> , <i>Withania somnifera</i> , <i>Solanum glabratum</i> , <i>Adenium obesum</i> , <i>Pistacia vera oleoresin</i> , <i>Caralluma quadrangula</i> , <i>Eulophia petersii</i> , <i>Phragmanthera austroarabica</i> , and <i>Asparagus officinalis</i> (Almehdar <i>et al.</i> , 2011), Terpenoids(Kuttan <i>et al.</i> , 2011), <i>Salvia officinalis</i> L. (sage) essential oil(Sertel <i>et al.</i> , 2011) Xanthone V(1) and 2-acetylfuro-1,4-naphthoquinone(Kuete <i>et al.</i> , 2011), Pomegranate extract(Nair <i>et al.</i> , 2011), Tea polyphenols(Singh M <i>et al.</i> , 2011; Chen D <i>et al.</i> , 2011), green tea extract (Chen D <i>et al.</i> , 2011; Cross <i>et al.</i> , 2011; Vu <i>et al.</i> , 2010; Lopez-Lazaro <i>et al.</i> , 2011; Liu X <i>et al.</i> , 2011), catechins from green tea(Shimizu <i>et al.</i> , 2011), methanolic leaf extract of <i>Indigofera cassioides</i> (MEIC)(Kumar <i>et al.</i> , 2011), phenolic compound from the wood of <i>Millettia leucantha</i> (Rayanil <i>et al.</i> , 2011), triterpenoid from the leaves of <i>Sinojackia sarcocarpa</i> (Wang O <i>et al.</i> , 2011), Phytochemicals green tea polyphenols (epigallocatechin gallate)(Wahl <i>et al.</i> , 2011), isoflavins from soy bean(Szliszka <i>et al.</i> , 2011; Jung <i>et al.</i> , 2011), biflavonoid amentoflavone(Lee S <i>et al.</i> , 2011), flesh of avocado fruits(D'Ambrosio <i>et al.</i> , 2011), polyphenolic compounds isolated from the leaves of <i>Leucenia leucocephala</i> (Haggag <i>et al.</i> , 2011), extracts of <i>Xanthium strumarium</i> (Cocklebur)(Takeda <i>et al.</i> , 2011), extracts from root bark of <i>Juglans Regia</i> L. (RBJR) (Hasan <i>et al.</i> , 2011), red beetroot (<i>Beta vulgaris</i> L.) extract(Kapadia <i>et al.</i> , 2011), Moringa oleifera leaf extract(Sreelatha <i>et al.</i> , 2011), Solvent extracts from the aerial and root parts and seed oil from E. sativa (rocket salad)(Khoobchandani <i>et al.</i> , 2011), <i>Toona sinensis</i> (leaf extracts)(Hseu <i>et al.</i> , 2011), water extract from <i>Mahonia bealei</i> (Fort.) Carr. Leaves (Hu <i>et al.</i> , 2011), Palm tocotrienols(Selvaduray <i>et al.</i> , 2010), Gugulipid (GL), extract from medicinal plant <i>Commiphora mukul</i> (Xiao D <i>et al.</i> , 2011), Methanol extracts of leaves of <i>Alnus sieboldiana</i> (Ludwiczuk <i>et al.</i> , 2011), ginger (<i>Zingiber officinale Roscoe</i>) (Tuntiwechapikul <i>et al.</i> , 2010), extract from black rice(Hui <i>et al.</i> , 2010), Cacao (<i>Theobroma cacao</i> L.)(Preza <i>et al.</i> , 2010), Periwinkle (<i>Catharanthus roseus</i>)(Liscombe <i>et al.</i> , 2010), <i>Panax stipuleanatus</i> rhizomes(Liang <i>et al.</i> , 2010), root extract of <i>Polygala senega</i> (Paul <i>et al.</i> , 2010), Black pepper (<i>Piper nigrum</i>) (Liu Y <i>et al.</i> , 2010), organic extracts of mulberry (<i>Morus alba</i> L.) leaves(Naowaratwattana <i>et al.</i> , 2010), <i>Phyllanthus emblica</i> L.(Ngamkitidechakul <i>et al.</i> , 2010), dry olive leaf extract(Mijatovic <i>et al.</i> , 2011), Cedrus deodara lignins(Saxena <i>et al.</i> , 2010).

(Chen C *et al.*, 2011). It is known to induce DNA damage and cause S and G2/M arrest in the cell cycle (Lu JJ *et al.*, 2012).

In breast cancer, curcumin is reported to prevent tumor growth by causing cell cycle arrest by inhibiting cyclin-dependent kinase (cdk) activity, suppressing cyclin D1 and cyclin E expression (Mukhopadhyay *et al.*, 2002), increasing levels of cdk inhibitors p21 and p27, inducing p53 transcriptional activity (Aggarwal *et al.*, 2007; Sen *et al.*, 2011), inhibition of matrix metalloproteinase-3 secretion (Boonrao *et al.*, 2010) and induce apoptosis by induction of Bax pathway (Choudhuri *et al.*, 2002). It is also reported to cause DNA damage and apoptosis in association with increased expression, phosphorylation, and cytoplasmic retention of the Breast cancer protein BRCA1 protein- a tumor suppressor protein which is a critical mediator of DNA repair in response to double-strand breaks (Rowe *et al.*, 2009).

In bladder cancer, curcumin has been reported to prevent tumor cell growth and induces apoptosis (Saini *et al.*, 2011) via decreased expression of the proapoptotic protein survivin and the angiogenic proteins vascular endothelial growth factor (VEGF) and VEGF receptor 1 (VEGFR1) (Chadalapaka *et al.*, 2008). In Acute Myeloid Leukemia (AML), curcumin promotes apoptosis and inhibit cell proliferation (Rao *et al.*, 2011) and is reported to inhibit telomerase activity in human leukemia cell HL-60 (Mukherjee *et al.*, 2007). In skin cancer, curcumin inhibits tumor progression by inhibiting the mammalian target for rapamycin or mTOR pathway (Phillips *et al.*, 2011).

In gastric cancer, curcumin suppresses cancer cell proliferation and invasion via down-regulation of P 21 activated kinase 1(PAK1) activity and cyclin D1 expression (Cai *et al.*, 2009) and overcome MDR (Tang *et al.*, 2005) and inhibits proliferation by affecting the cell cycle (Moragoda *et al.*, 2001). In ovarian carcinoma curcumin has been reported to inhibit tumor growth and

angiogenesis via nuclear factor-kappa B pathway (Lin *et al.*, 2007). Curcumin has been reported to induce apoptosis in nasopharyngeal cancers by activation of Reactive Oxygen Species (ROS), mitochondrial depolarization and caspase 3 dependant pathways and prevents tumor cell growth and alters the phenotype of migratory cells (Kuo *et al.*, 2011; Wang *et al.*, 2011a; Wang *et al.*, 2011b; Wong *et al.*, 2010).

In liver cancer, curcumin has been reported to cause cell death and promote mitochondria mediated apoptosis (Qian *et al.*, 2011) and inhibits tumor cell growth (Cheng *et al.*, 2010; Ning *et al.*, 2009). Curcumin has been reported to induce apoptosis in pancreatic cancer (Sahu *et al.*, 2009). to inhibit matrix metalloproteinase protein-2, MMP-2 in human laryngeal squamous carcinoma cells (Mitra *et al.*, 2006) promote apoptosis in human lung adenocarcinoma cells (Zhang J *et al.*, 2010) by DNA damage (Saha *et al.*, 2010), caspase pathways and ER Stress mechanisms (Wu *et al.*, 2010). Curcumin has been reported to inhibit growth of uterine (Tsuiji *et al.*, 2011), ovarian cancer (Saydmohammed *et al.*, 2010; Watson *et al.*, 2010; Seo *et al.*, 2010), brain and nervous system cancer (Zanotto-Filho *et al.*, 2011; Spiller *et al.*, 2011). It is known to cause apoptosis and prevent cell growth in Hodgkins lymphoma (Mackenzie *et al.*, 2008), lymphoma (Vishvakarma *et al.*, 2011; Zhang *et al.*, 2010; Li *et al.*, 2009; Xiao *et al.*, 2010; Zhongguo *et al.*, 2008) and esophageal cancers (O'Sullivan-Coyne *et al.*, 2009; Tian *et al.*, 2008). It is reported to participate in mechanisms aiding in overcoming MDR in multiple myeloma (Xiao *et al.*, 2010). The overall effect of curcumin in different cancer has been summarized in **Table-2**.

Curcumin formulations and recent developments

Although curcumin offers promise to cancer research over the conventional toxic chemotherapeutic drugs, the main disadvantage of applicability of curcumin in disease therapy is its lipophilic nature and poor aqueous solubility, minimum bioavailability and



Table 2: Major effects of Curcumin and its derivatives in different Cancers

Role of curcumin	Target molecules/pathways		Cancer
	Direct	Indirect	
Overcome drug resistance	ABC transporter TLR4 dimerization I κ B α kinase Cyclooxygenase-2 Protein kinase C Protein kinase A Phosphorylase kinase	p53-p300 cross-talk leading to cell death pathway	Breast cancer(Sen <i>et al.</i> , 2011)
		Proteasome pathway	Multiple myeloma(Mujtaba <i>et al.</i> , 2012)
		Down-regulating the activity of NF- κ B	Gastric cancer cells (Tang <i>et al.</i> , 2005; Yu LL <i>et al.</i> , 2011)
Epigenetic	pp60c-src tyrosine kinase Ca ²⁺ -dependent protein kinase Xanthine oxidase Ca ²⁺ -ATPase of sarcoplasmic reticulum	Decrease of pro-caspase 3 pro-caspase 9, increase of PARP cleavage and the ratio of Bax/Bcl-xL	K562/A02(Lu JJ <i>et al.</i> , 2012)
		Demethylation of the Neurog1 gene and restored the expression of this cancer-related CpG-methylation epigenome marker gene	Prostate cancer(Shu <i>et al.</i> , 2011)
Apoptosis	Inositol 1,4,5-triphosphate receptor (Shrikanth <i>et al.</i> ,1994)	Down Regulation of histone deacetylases, histone acetyltransferases, DNA methyltransferase I, and miRNAs	Cancer(Reuter <i>et al.</i> , 2011)
		Activation of Apoptosis pathways	Prostate cancer(Yallapu <i>et al.</i> , 2011b)
		Induce apoptosis by decreased phosphorylation of Retinoblastoma (Rb).	Pituitary tumour(Schaaf <i>et al.</i> , 2010)
		Activation of ROS pathway, DNA damage mitochondria-mediated and ER stress-dependent pathways.	Myelomonocytic leukemia (Huang <i>et al.</i> , 2011)
		Down Regulation of caspase cascade	Breast cancer cell line(Zong <i>et al.</i> , 2011)
		Down-regulation of Bcl-2 and procaspase-3 and increased production of reactive oxygen species (ROS) level	Adenocarcinoma cell line (Ibrahim <i>et al.</i> , 2008)
		Increasing Bax expression, decreasing the expression of Bcl-2 and Bcl-xL, decreasing mitochondrial membrane potential, increased ROS	Small Cell Lung Cancer (Yang <i>et al.</i> , 2011)
		ROS level Up regulation of Bax and down regulation of Bcl-2, mitochondrial dysfunction	Human nasopharyngeal carcinoma cells (Kuo <i>et al.</i> , 2011)
		Bcr-Abl suppression	Chronic myeloid leukemia (Acharya <i>et al.</i> , 2011)
		Down regulation of NF- κ B expression	Oesophageal cancer (Tian <i>et al.</i> , 2008)
Inhibits invasion and metastasis	Proapoptotic via cytotoxicity of Cum-np	FAS/caspase-8 (extrinsic) pathway and ER stress proteins, growth arrest- and DNA damage-inducible gene 153 (GADD153) and glucose-regulated protein 78 (GRP78)	Human non-small cell lung cancer cells (Wu <i>et al.</i> , 2010)
		Induction of apoptosis by cleavage of PARP, caspase-3, and reduction in Bcl-XL levels	Gastric and colon cancer (Moragoda <i>et al.</i> , 2001)
		Down-regulating the NF- κ B transcription factor	Human mammary epithelial carcinoma MCF-7 cells (Zong <i>et al.</i> , 2011)
Tumor suppression	Induces a tumor-suppressive miRNA, miR-203	Down regulation of Matrix metalloproteinase-3 (MMP-3)	Human invasive breast carcinoma cells (Yallapu <i>et al.</i> , 2010b)
		Reduces SCC-4 cell invasion, leads to the recruitment of alpha-tubulin.	Bladder cancer (Rejinold <i>et al.</i> , 2011c)
		Inhibits tumor growth by inhibiting angiogenesis	Human tongue squamous cell carcinoma (Chen JW <i>et al.</i> , 2011)
		Inhibition of STAT3 signaling pathway via LLL12 and FLLL32, down regulation of cyclin D1, Bcl-xL	Colon cancer (Gou <i>et al.</i> , 2011)
		Inhibits tumor cell proliferation	Human rhabdomyosarcoma cells (Wei <i>et al.</i> , 2011)
		Inhibition of Activation of nuclear factor-kappaB	Jurkat cell lines (Yadav <i>et al.</i> , 2010)
		Disturbed mitotic spindle structure, activated mitotic check points	Pancreatic cancer (Bisht <i>et al.</i> , 2010)
		G1/S arrest	MCF-7 cells (Banerjee <i>et al.</i> , 2010)
	Inhibition of NF- κ B signaling		Drug resistant AML cell lines (Rao <i>et al.</i> , 2011)
			Medulloblastoma (Spiller <i>et al.</i> , 2011)

targeted delivery to the transformed cell.

Different formulations and delivery devices are being tested to enable curcumin as an effective agent in cancer management. Biocompatible urithin polymers (PU) from polylactic acids and hexamethylene diamide (Selvaraj *et al.*, 2011), liposomes coated with N-trimethyl chitosan chloride (TMC) (Chen H *et al.*, 2011) have been reported to function as efficient formulations. Cationic and anionic curcumin conjugates by anchoring curcumin (Cur) onto poly (vinylpyrrolidone) (PVP-Cur) and onto hyaluronic acid (HA-Cur) (Manju *et al.*, 2011b) are also being exploited. Delivery through formulations like niosomes (Rungphanichkul *et al.*, 2011), amphiphilic polymers like Lauroyl sulphated chitosan (LSCS) (Shelma *et al.*, 2011) and carboxy methyl derivatives (Anitha *et al.*, 2011), fibrinogen nanoparticles (CRC-FNPs) (Rejinold *et al.*, 2011a), nanospheres (Mukerjee *et al.*, 2009), 2-Hydroxypropyl- γ -cyclodextrin/curcumin-liposomal nanoparticles (Dhule *et al.*, 2011) have been reported to be effective. Nanoparticles (Li R *et al.*, 2011; Yallapu *et al.*, 2010a; Yallapu *et al.*, 2010b) reported for their biocompatible, non toxic, biodegradable nature and thermoresponsive properties (Rejinold *et al.*, 2011c) have shown positive results in curcumin delivery. Conjugated nanoparticle copolymers with Chitosan-g-poly (N-isopropylacrylamide) nanoparticles (Rejinold *et al.*, 2011b), amphiphilic methoxy polyethylene glycol-poly (caprolactone) (mPEG-PCL) are being employed (Shao *et al.*, 2011). Magnetic particles are being tested for controlled drug delivery (Koppolu *et al.*, 2010; Chin *et al.*, 2010; de-Souza *et al.*, 2011).

Strategies to increase the solubility of analogues of curcumin (Zhang *et al.*, 2011), lipid-based formulations (Thangapazham *et al.*, 2008; Yu *et al.*, 2011; Sethacheewakul *et al.*, 2011; Xie *et al.*, 2011), nanosuspensions (Zhang H *et al.*, 2011), liposomes (Chen H *et al.*, 2011; Dhule *et al.*, 2011; Li *et al.*, 2005; Mach *et al.*, 2009; Pandelidou *et al.*, 2011; Agashe *et al.*, 2011), microemulsions (Liu CH *et al.*, 2011), polymer encapsulations (Mohanty *et al.*, 2010; Sahu *et al.*, 2011;

Das *et al.*, 2010), aerosols (Selvam *et al.*, 2011), and nanodisks (Singh AT *et al.*, 2011; Tadmor *et al.*, 2011; Ghosh *et al.*, 2011) are being tested as delivery formulations of curcumin in different cancer.

Few reports on targeted delivery (Thamake *et al.*, 2011) of curcumin exists. Tissue specific targeting are being tested like curcumin loaded Eudragit® S100 coated calcium pectinate microsphere in colon cancer (Zhang L *et al.*, 2011). Reports on the use of Gelatin microspheres (C-GMS) in lung cancer (Cao *et al.*, 2011) and Gelucire44/14 in eyes (Liu R *et al.*, 2011) have shown promising results. The application of different formulations of curcumin in different cancer is summarized in **Table-3**.

DISCUSSIONS

Curcumin has shown strong reports in its anti-cancerous activities in different malignancies including brain, skin, lung, prostate, breast, ovarian, liver, nasopharyngeal, gastrointestinal, pancreatic and colorectal cancers. Synergistic effects of curcumin with agents like decetaxel has also been reported in lung cancer (Yin *et al.*, 2011). However, because of its both pro and anti-oxidant effect, it has been reported to behave like a “double edged sword”. (Kawanishi *et al.*, 2005) and its safety as a chemopreventive agent remains yet to be exploited. The future scope of this review remains in potential applications of targeted cancer cell killing by a natural product like curcumin.

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**Table 3: Curcumin and Formulations : Cancer Research**

Cancer	Delivery Systems
Lung adenocarcinoma H441 cells and nude rats bearing xenograft H441 tumors	Cyclodextrin entrapped curcuminoid derivative(Agashe <i>et al.</i> , 2011)
Skin cancer	Micro emulsions for transdermal delivery(Liu <i>et al.</i> , 2011)
Non small cell lung cancer	Intravenous synergistic with Docetaxel(Yin <i>et al.</i> , 2011)
Lung cancer cells	Matrix of Urethane polymers (PU) prepared from low-molecular weight polylactic acid (PLA) and hexamethylene diisocyanate (HMDI) curcumin-containing PU membranes(Selvaraj <i>et al.</i> , 2011)
Glioma cells and Caco-2 cells	Magnetic nanoparticles (MNPs) (Manju <i>et al.</i> , 2011a)
L929 (mouse fibroblast), PC3 (prostate) and MCF7 (breast) cancer cell lines	Fibrinogen nanoparticles (Rejinold <i>et al.</i> , 2011b)
Tumor mice model	Curcuminoids-loaded solid lipid nanoparticles (curcuminoids-SLNs) and curcumin-loaded solid lipid nanoparticles (curcumin-SLNs)(Li <i>et al.</i> , 2005)
Osteosarcoma	Liposomal nanoparticles(Dhule <i>et al.</i> , 2011)
Cancer cells	N,O-Carboxymethyl Chitosan Nanoparticles(Anitha <i>et al.</i> , 2011)
Bcr-Abl + leukemia cells	Nanoparticles(Acharya <i>et al.</i> , 2011)
PC3, L929 cells	Biocompatible thermoresponsive polymeric chitosan-g-poly (N-vinylcaprolactam) nanoparticles (TRC-NPs)(Rejinold <i>et al.</i> , 2011c)
Colorectal cell lines	Lyophilised egg PC liposomes (Pandelidou <i>et al.</i> , 2011)
C6 Glioma cells	Methoxy polyethylene glycol-poly (caprolactone) nanoparticles (Shao <i>et al.</i> , 2011)
3T3-L1 preadipocytes and adipocytes	Conjugated with Polyethylene glycol (Kim <i>et al.</i> , 2011)
Breast cells	Surface functionalized polymeric PLGA nanoparticles by non-covalent insertion of a homo-bifunctional chemical crosslinker, bis(sulfosuccinimidyl) suberate (BS3) for targeted cancer therapy (Thamake <i>et al.</i> , 2011)
L929 cells	Hollow microcapsules (Manju <i>et al.</i> , 2011 b)
Cisplatin resistant A2780CP ovarian and metastatic MDA-MB-231 breast cancer cells	Encapsulated PGLA formulation (Yallapu <i>et al.</i> , 2010a)
Prostrate cancer	Poly(β -cyclodextrin)/curcumin self-assembly (Yallapu <i>et al.</i> , 2010b), Nanoparticles (Thangapazham <i>et al.</i> , 2008), cellulose nanoparticles (Yallapu <i>et al.</i> , 2011a), PGLA nanospheres (Mukerjee <i>et al.</i> , 2009)
Sub cutaneous injection in mice	A biodegradable and biocompatible polymer, poly (d,l-lactide-co-glycolide), was used to fabricate curcumin microparticles (Shahani <i>et al.</i> , 2010)
Pancreatic cell lines MIA PaCa-2 and PANC-1	Encapsulated methoxy poly (ethylene glycol) (MePEG)/poly-epsilon-caprolactone (PCL) diblock copolymeric micelle (Mohanty <i>et al.</i> , 2010)
Hela cells	Micelles of Pluronic encapsulated curcumin (Sahu <i>et al.</i> , 2011), Alginate-chitosan-pluronic composite nanoparticles (Das <i>et al.</i> , 2010)
Pancreatic cancer	Liposome mediated (Li <i>et al.</i> , 2005)

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