Journal of Research in Biology



OPEN ACCESS

Original Research Paper

An International Online Open Access Publication group

Effect of *Ulva lactuca* extract on growth and proximate composition of *Vigna unguiculata* l. Walp.

Authors:

Gireesh R¹⁺, Haridevi CK² and Salikutty Joseph³

Institution:

^{1, 3}Department of Horticulture, Kerala Agricultural University, Vellanikkara P.O., Thrissur, Kerala, India 680656.

²National Institute of Oceanography, RC- Kochi, Ernakulam North P.O., Ernakulam, Kerala, India 682018.

⁺ Present address: Central Marine Fisheries Research Institute, North P.O., Ernakulam, Kerala, India 682018.

As organic farming gains more attention, seaweed cultivation and its utilization may be an economical approach in agricultural production. The effect of seaweed *Ulva lactuca* extract on growth and proximate composition of *Vigna unguiculata* L. Walp was studied. Seedlings of *V. unguiculata* soaked with seaweed extract performed better when compared to seedlings with water soaked control. The low concentration (20%) of aqueous seaweed extract promoted seedling growth in terms of shoot length, root length, fresh and dry weight and chlorophyll and carotenoids content. The biochemical constitutents show similar patterns with protein content of shoot and root, amino acid of shoot and root, α -amylase and β -amylase activities being higher with seaweed extract in *V. unguiculata*. The seaweed extract in these experiments showed as biological fertilizer, which is a nontoxic and ecofriendly supplement to chemical fertilizer for many crops intended to attain higher yields.

Keywords:

ABSTRACT:

Crop, seaweed extract, biochemical composition, fertilizer, organic farming

Corresponding author: Gireesh R

girmsr@gmail.com

Article Citation: Gireesh R, Haridevi CK and Salikutty Joseph Effect of *Ulva lactuca* extract on growth and proximate composition of *Vigna unguiculata* I. Walp. Journal of research in Biology (2011) 8: 624-630

Dates:

Received: 17 Nov 2011 /Accepted: 26 Nov 2011 /Published: 13 Dec 2011

Phone No: 91 9496446725

Email:

© Ficus Publishers.

This Open Access article is governed by the Creative Commons Attribution License (http:// creativecommons.org/licenses/by/2.0), which gives permission for unrestricted use, non-commercial, distribution, and reproduction in all medium, provided the original work is properly cited.

Web Address: http://jresearchbiology.com/ Documents/RA0148.pdf.

Journal of Research in biology

An International Open Access Online Research Journal Submit Your Manuscript www.ficuspublishers.com

624-630 | JRB | 2011 | Vol 1 | No 8

www.jresearchbiology.com



INTRODUCTION

Seaweeds have been used as green manure, cattle feed, food for human consumption and as a source of phycocolloids such as sugar, alginic acid and carrageenan (Chapman, 1970). Besides their application as green manure, liquid extracts obtained from seaweeds (SLF) have gained importance as foliar sprays for several crops (Thivy, 1961, Bokil et al, 1974) because the extract contains growth promoting hormones auxins (IAA. IBA), cytokinins, trace elements, vitamins and amino acids (Challen and Hemingway 1965). Thus, these extract when applied to seeds or when added to the soil, can stimulate growth of plants (Blunden, 1971). Green manure was found to be better than chemical fertilizers because of the high level of organic matter and colloidal compounds that aids in retaining soil moisture and minerals in the upper soil level that are available to the roots (Wallen, 1955). Booth (1969) observed that the value of seaweeds as fertilizers was not only due to nitrogen, phosphorus and potash content, but also because of the presence of trace elements and metabolites. Aqueous extract of Sargassum wightii when applied as a foliar spray on Zizyphus mauritiana showed an increase yield and quality of fruits (Ramarao, 1991). Seaweed extracts are now available commercially as Maxicrop, Algifert Marinure, Goemar GA14, Kelpak 66, Seaspray, Cytex, Seasol and Seacrop 16. Recent, research demonstrated that seaweed fertilizers can compete with other fertilizers and are very economical (Gandhiyappan and Perumal, 2001). This study was undertaken to investigate the effect of seaweed liquid extract on the growth and biochemical characteristics of Vigna unguiculata.

MATERIALS AND METHODS

Seaweed Ulva lactuca (Chlorophyceae) used in the present study was collected from the coastal area of Rameswaram, India (9°25' N and 79°15' E). The algal species were washed thoroughly with seawater to remove all unwanted impurities, adhering sand particles and epiphytes. Morphologically distinct thallus of algae were placed separately in new polythene bags and were kept in an ice box containing slush ice and transported to the laboratory. Samples were washed thoroughly using tap water to remove the salt on thallus surface. The water was drained off and the algae were spread on blotting paper to remove excess water. One kg of seaweed was cut into small pieces, boiled with one litre of distilled water for an

hour, and filtered through GF/C paper. The filtrate was taken as 100% concentration of the seaweed extract and from this stock solution, different concentrations (5%, 10%, 20%, 30%, 40% and 50%) were prepared using distilled water (Bhosle et al, 1975). As the seaweed liquid fertilizers contained organic matter, they were refrigerated between 0 and 4 °C. The colour, pH, calcium, magnesium, sodium, potassium, iron, chloride, sulphur, silicon, aluminium, zinc, copper and nitrate content were analyzed by the method described by American Public Health Association (APHA, 1995).

The crop plant selected for the present study was *Vigna unguiculata* belonging to the family Fabaceae. Seeds with uniform size, colour and weight obtained from Seed Bank of Kerala Agricultural University chosen for the experimental purpose.

Experiment I (water soaked)

Eight hundred uniform sized seeds were soaked in water for 24 hours. After soaking, they were divided into batches of 100 seeds each and were placed in 8 Petri dishes with filter paper. One batch of seeds was considered as the control and they were watered with 10 ml of tap water every 24 hours. The reminder of batches were treated once with 10 ml/Petri dish of 5%, 10%, 20%, 30%, 40% and 50% of aqueous seaweed extract every 24 hours.

Experiment II (seaweed extract soaked)

One hundred seeds were soaked for each concentration of aqueous seaweed extracts for 24 hours and then were placed in various Petri dish plates with filter paper and sprayed water regularly every 24 hours. Water soaked seeds were used as controls. Samples were taken from each set 15 days after sowing. The growth parameters including germination percentage, fresh and dry weight and shoot and root length was calculated. Seven biochemical constituents, chlorophyll content (Arnon, 1949), carotenoid (Mackinney, 1941), protein (Lowry et al, 1951), amino acids (Moore and Stein 1948), total sugar content (Nelson, 1944) and α -amylase and β -amylase activities (Bernfeld, 1955) were estimated from V. unguiculata seedlings. Data were statistically analyzed using correlation of coefficient method and all the measurements were made triplicate.

RESULTS

The physico-chemical properties of seaweed *Ulva lactuca* extract was analyzed (**Table I**). The

Ulva lactuca extract								
Parameters	Seaweed extract							
Physical parameters								
Colour	Yellow							
pH	7.3							
Chemical parameters (mg.l)								
Sodium	185.00							
Potassium	113.00							
Magnesium	108.30							
Calcium	195.26							
Phosphorous	51.35							
Iron	0.37							
Chloride	415.55							
Sulphate	16.84							
Silica	38.12							
Copper	0.38							
Zinc	1.01							
Nitrate	19.05							

Table I. Physico-chemical properties of seaweedUlva lactuca extract

pH of the yellow coloured extract was 7.3 and had high levels of calcium, sodium, potassium, magnesium, phosphorous, iron and chloride. The effect of extract on germination percentage and growth of *V. unguiculata* are presented in **Tables IIa and IIb.** Maximum seed germination (99%) was found at 20% concentration in both water

soaked and seaweed extracts soaked seeds. The germination percentage increased with concentration up to 20% and thereafter it declined. No germination was seen at concentrations above 50%. The lowest germination percentage (19%) was found in water soaked seeds at 50% concentration. The highest shoot length (15.7 cm/ seedling), root length (5.41 cm/seedling), fresh and dry weight (3.970, 0.807 g/seedling) was observed at 20% concentration of seaweed extract soaked plants. The lowest shoot length (8.20 cm/seedling), fresh and dry weight (1.980, 0.701 g/seedling) were found at 50% concentration of seaweed extract soaked seeds The biochemical constituents increased with concentration levels up to 20% and thereafter declined (Tables III a-c). The highest values of chlorophyll content (2.268 mg/g fr. Wt.), carotenoid (0.852 mg/g fr. wt.), amino acid content of shoot (1.343 mg/g fr. wt.), α and β amylase (1.639, 1.613 µg/min/mg protein respectively), total sugar content of shoot and root (11.207, 8.602 mg/g fr. wt.) were recorded at 20% seaweed extract soaked seedlings. The lowest values were observed at 50% U. lactuca extract soaked seedlings (Tables III a-c). There was a significant difference in growth and biochemical status at different

 Table IIa. Effects of seaweed extract on germination and growth of V. unguiculata seedlings

	Germination (%)		Shoot length (cm/seedling)	Root length (cm/seedling)		
	Ι	II	Ι	II	Ι	П	
Control	85 ± 2.65	85 ± 2.65	7.0 ± 0.20	7.0 ± 0.20	2.5 ± 0.06	2.5 ± 0.06	
5%	93 ± 0.58	97 ± 0.58	10.7 ± 0.20	11.7 ± 0.06	2.6 ± 0.12	2.6 ± 0.06	
10%	94 ± 1.0	97 ± 1.0	13.3 ± 0.15	13.9 ± 0.06	3.0 ± 0.12	3.5 ± 0.12	
20%	99 ± 0.58	99 ± 0.58	14.2 ± 0.06	15.7 ± 0.21	4.2 ± 0.06	5.4 ± 0.06	
30%	91 ± 1.7	93 ± 0.58	12.4 ± 0.06	13.1 ± 0.20	4.4 ± 0.10	4.8 ± 0.06	
40%	65 ± 0.58	78 ± 1.00	8.60 ± 0.10	9.3 ± 0.20	3.0 ± 0.12	4.6 ± 0.07	
50%	19 ± 1.15	37 ± 11.0	8.20 ± 1.15	8.7 ± 0.06	2.9 ± 0.05	2.9 ± 0.20	

I - Water soaked; II- Seaweed extract soaked

Table IIb. Effects of seaweed extract on germination and growth of V. unguiculata seedlings

Seedling (g/seedling)							
	Fresh weight		Dry weight				
	I II		Ι	II			
Control	1.82 ± 0.06	1.82 ± 0.06	0.679 ± 0.00	0.679 ± 0.00			
5%	1.99 ± 0.58	2.06 ± 0.12	0.719 ± 0.01	0.731 ± 0.00			
10%	2.07 ± 0.6	3.93 ± 0.05	0.782 ± 0.00	0.873 ± 0.01			
20%	2.52 ± 0.03	3.97 ± 0.04	0.807 ± 0.01	0.882 ± 0.01			
30%	2.12 ± 0.01	3.73 ± 0.36	0.800 ± 0.00	0.875 ± 0.01			
40%	1.78 ± 0.02	1.98 ± 0.01	0.701 ± 0.15	0.790 ± 0.00			
50%	1.85 ± 0.05	1.98 ± 0.01	0.688 ± 0.01	0.701 ± 0.01			

I – Water soaked; II- Seaweed extract soaked





	Leaf (mg/g fr. wt.)							
	Chlor	ophyll	Carot	enoids				
	Ι	II	Ι	II				
Control	1.253 ± 0.11	1.253 ± 0.11	0.667 ± 0.01	0.667 ± 0.01				
5%	1.293 ± 0.02	1.793 ± 0.02	0.790 ± 0.00	0.826 ± 0.01				
10%	1.370 ± 1.01	1.960 ± 0.00	0.826 ± 0.00	0.840 ± 0.01				
20%	1.427 ± 0.01	2.268 ± 0.02	0.850 ± 0.00	0.852 ± 0.00				
30%	1.110 ± 0.00	1.226 ± 0.00	0.604 ± 0.00	0.647 ± 0.02				
40%	0.980 ± 0.01	1.000 ± 0.00	0.355 ± 0.01	0.422 ± 0.01				
50%	0.595 ± 0.00	0.624 ± 0.01	0.310 ± 0.01	0.711 ± 0.01				
		Leaf (mg/g fr. wt.)						
	α-an	nylase	β-an	nylase				
	Ι	II	Ι	II				
Control	1.388 ± 0.003	1.388 ± 0.003	1.354 ± 0.000	1.354 ± 0.000				
5%	1.402 ± 0.001	1.522 ± 0.020	0.378 ± 0.001	1.404 ± 0.010				
10%	1.477 ± 1.064	1.530 ± 0.001	0.383 ± 0.001	1.466 ± 0.010				
20%	1.636 ± 0.005	1.639 ± 0.010	1.566 ± 0.005	1.613 ± 0.031				
30%	0.852 ± 0.045	0.896 ± 0.010	1.125 ± 0.576	0.864 ± 0.010				
40%	0.396 ± 0.003	0.509 ± 0.031	0.668 ± 0.005	0.765 ± 0.011				
50%	0.289 ± 0.003	0.288 ± 0.001	0.606 ± 0.005	0.734 ± 0.030				

I – Water soaked; II- Seaweed extract soaked

	Shoot (mg/g fr. wt.)								
	Total sugar		Pro	otein	Amino acid				
	Ι	II	Ι	II	Ι	II			
Control	8.340 ± 0.003	8.340 ± 0.003	1.548 ± 0.001	1.548 ± 0.001	0.434 ± 0.001	0.434 ± 0.005			
5%	10.004 ± 0.01	10.280 ± 0.02	2.513 ± 0.002	2.632 ± 0.004	0.627 ± 0.003	0.646 ± 0.003			
10%	10.632 ± 0.003	10.670 ± 0.004	2.915 ± 0.004	3.084 ± 0.004	0.847 ± 0.003	0.864 ± 0.005			
20%	11.270 ± 0.003	11.290 ± 0.016	3.156 ± 0.005	3.338 ± 0.005	1.279 ± 0.002	1.343 ± 0.002			
30%	8.276 ± 0.002	8.340 ± 0.002	1.315 ± 0.004	1.503 ± 0.012	0.787 ± 0.002	0.858 ± 0.001			
40%	6.369 ± 0.010	6.391 ± 0.006	0.983 ± 0.001	1.114 ± 0.040	0.691 ± 0.001	0.752 ± 0.001			
50%	0.509 ± 0.008	0.553 ± 0.003	0.863 ± 0.006	0.983 ± 0.007	0.602 ± 0.002	0.616 ± 0.001			

I - Water soaked; II- Seaweed extract soaked

Table IIIc. Effects of seaweed extract on proximate composition of V. unguiculata root

			Root (mg/g fr. v	vt.)			
	Total	sugar	Protein		Amino acid		
	Ι	II	Ι	II	Ι	II	
Control	5.803 ± 0.003	8.340 ± 0.003	1.124 ± 0.005	1.124 ± 0.005	0.407 ± 0.003	0.407 ± 0.003	
5%	7.481 ± 0.002	7.533 ± 0.010	2.124 ± 0.004	2.120 ± 0.056	0.430 ± 0.002	0.480 ± 0.010	
10%	7.816 ± 0.003	7.906 ± 0.005	2.131 ± 0.002	2.142 ± 0.002	0.493 ± 0.006	0.554 ± 0.004	
20%	8.496 ± 0.001	8.602 ± 0.001	2.542 ± 0.008	2.721 ± 0.008	0.554 ± 0.003	0.684 ± 0.010	
30%	6.433 ± 0.003	6.627 ± 0.037	0.551 ± 0.026	0.555 ± 0.003	0.506 ± 0.006	0.633 ± 0.010	
40%	4.394 ± 0.007	4.620 ± 0.011	0.429 ± 0.001	0.452 ± 0.016	0.451 ± 0.002	0.595 ± 0.008	
50%	3.492 ± 0.003	3.001 ± 0.021	0.400 ± 0.003	0.413 ± 0.003	0.408 ± 0.002	0.440 ± 0.002	

I - Water soaked; II- Seaweed extract soaked



concentration levels. Correlation of coefficient was carried out to find the significance level (*p<0.05 and ** p<0.01). High significance was observed in 20% concentration (**Table IV**).

DISCUSSION

Vigna unguiculata seeds soaked with lower concentration (5-40%) of the seaweed extract showed high germination rates (85-95%), while higher concentrations (\geq 40%) have inhibited germination. The increased seedling growth may be due to the presence of phenyl acetic acid and other closely related compounds in the extract (Taylor and Wilkinson, 1977) as well as the presence of growth promoting hormones like auxins, gibberellins, cytokinins, trace elements, vitamins and amino acids (Challen and Hemingway, 1965). The present findings agree with field trials in other crops such as in *Cajanus cajan* (Mohan *et al.*,

1994), maize, ragi and kambu (Rajkumar and Subramanian. 1999) and *Dolichos* buflorus (Anantharaj and Venkatesalu, 2002). Statistically significant differences were observed for leaf pigments and total sugar contents in both shoot and root. A positive response was observed for shoot length at 10-20% seaweed extract soaked seedlings. Trace elements, especially calcium that exists in this seaweed extract are in a naturally chelated form, which can absorb more readily than from soil. The higher concentrations ($\geq 40\%$) showed a decreasing for V. unguiculata. Similar results were recorded in C. cajan (Mohan et al., 1994) and Vigna radiata (Venkataraman et al., 1993) where maximum seedling growth occurred at lower concentrations of Padina extracts. Dhargalkar and Untawale (1983) also reported comparable results with red and brown algal extracts on the growth of chillies, turnips and pineapple.

			5%	10%	20%	30%	40%	50%
Seed	Germination	Ι	0.655	0.866*	1.000*	0.500	-1.000	-0.500
		II	0.655	0.866*	0.500	0.500	-0.866	0.419
Seedling	Fresh weight	Ι	-0.904	-0.999	-0.533	-0.999	-0.914	-0.983
		II	-0.604	-0.785	-0.997	-1.000	-0.821	-0.569
Seedling	Dry weight	Ι	-0.329	0.245	-0.456	-0.381	0.381	-0.381
		II	-0.520	-0.381	0.536	0.610	-0.893	-0.810
Shoot	Length	Ι	0.500	-0.327	-0.866	-0.866	0.500	-
		II	-0.866	0.866*	0.961**	0.500	0.500	-0.866
Root	Length	Ι	-0.500	0.500	-0.500	-0.500	-1.000	-0.500
		II	-0.866	-0.500	-0.500	-1.000	-0.500	-
Leaf	Chlorophyll	Ι	-0.987	-0.172	-0.294	-0.939	-0.980	-0.985
		II	-0.987	-0.857	0.965**	0.284	-1.000	0.054
	Carotenoids	Ι	0.189	-0.500	0.933*	-0.156	-	-0.286
		II	0.292	0.768*	0.933*	0.115	0.143	0.069
	α - amylase	Ι	-0.993	-0.915	0.999**	-0.803	-0.300	-0.845
		II	0.999**	0.981**	0.995**	-0.826	-0.767	-0.596
	β - amylase	Ι	-0.189	-0.655	0.945*	-0.945	-0.968	-0.866
		II	-0.945	-0.904	0.811*	-0.918	-0.945	-0.951
Shoot	Total sugar	Ι	-0.746	0.700	0.700	-0.803	0.684	0.545
		II	-0.797	0.717	0.765*	-0.127	0.115	0.500
	Protein	Ι	0.655	0.500	0.101	0.500	0.500	0.545
		II	0.427	0.277	0.819	0.126	0.062	0.352
	Amino acid	Ι	-0.882	-	-1.000	-0.891	-0.327	-0.756
		II	0.945	0.945*	-0.786	-0.945	-0.655	-0.189
Root	Total sugar	Ι	-0.169	0.300	0.945*	0.655	0.655	_
		II	0.933*	0.929*	0.982**	-0.644	-0.810	-0.304
	Protein	Ι	-0.349	-0.023	0.928	0.658	-0.600	-0.911
		II	-0.994	0.877*	-0.890	-0.120	-0.970	0.392
	Amino acid	Ι	-0.091	-0.583	0.529	-0.137	0.500	0.371
		II	-0.374	0.327	-0.693	-0.176	0.047	-0.619

I – Water soaked; II- Seaweed extract soaked Significant at * p<0.05 level; Significant **p< 0.01 level



The lower concentrations of the extract also promoted the chlorophyll content of *V. unguiculata* up to 20% when compared to the control while higher concentrations (> 20%) decreased the chlorophyll content. A similar observation was made in *Vigna mungo* (Venkataraman and Mohan, 1997) and when seaweed extract (15-20%) applied as foliar spray enhanced the leaf chlorophyll level in plants (Blunden *et al.*, 1996).

The highest total sugar content was recorded at 20% concentration of seaweed extract soaked treatment in V. unguiculata. The sugar content increased up to 20% concentration of seaweed extract and decreased at higher concentrations. The increase in the total sugar content at lower concentration of seaweed extract might be due to absorption of most of the necessary elements (Kannan and Tamilselvan, 1990). The same trend was observed in the H. musciformis with NPK application in black gram (Tamilselvan and Kannan, 1994), and D. biflorus (Anantharaj and Venkatesalu, 2002). It has been observed that α – amylase activity was higher than the β – amylase activity. Both α – amylase and β -amylase activity increased at lower concentrations and decreased in higher concentrations. In conclusion, Ulva lactuca extract was found to be a promising fertilizer. The extract act as a biological fertilizer for organic farming, which is non-toxic, non-flammable for attaining better seed germination and growth.

ACKNOWLEDGMENT

The authors are grateful to the Head, Department of Olericulture, Kerala Agricultural University, Thrissur, Kerala, India for the facilities provided.

REFERENCES

Anantharaj M and Venkatesalu V. 2002. Studies on the effect of seaweed extracts on *Dolichos biflorus*. Seaweed Research and Utilization 24 (1):129-137.

APHA. 1995. Standard methods for examination of water and wastewater analysis, 19th ed., APHA, Washington. 231.

Arnon DI. 1949. Copper enzymes in isolated chloroplasts, polyphenol oxydase in *Beta vulgaris*. Plant physiology 12:1-15.

Bernfeld P. 1955. Amylase α and β . In: Methods in Enzymology. Vol 1. (Colowick, SP and Kaplan ND

(Eds.). Academic Press Inc., New York. 149-150.

Bhosl NB, Untawale AG and Dhargalker VK. 1975. Effects of seaweed extract on growth of *Phaseolus vulgaris*. Indian Journal of Marine Sciences 4:208-210.

Blunden G. 1971. The effect of aqueous seaweed extract as fertilizer additives. Proceedings of Sixth International Seaweed Symposium, Tokyo, (Blunden G (Ed.), 584-589.

Blunden G, Jenkins T and Liu YW. 1996. Enhanced chlorophyll levels in plants treated with seaweed extract. Journal of Applied Phycology 8:535-543.

Bokil KK, Mehta VC and Datat DS. 1974. Seaweed as manure: II pot culture manurial experiments on wheat. Phykos 13:1-15.

Booth E. 1969. The manufacture and properties of liquid seaweed extracts. Proceedings of Sixth International Seaweed Symposium, Tokyo, (Blunden G (Ed.), 655-662.

Challen SB and Heminway JC. 1965. Growth of higher plants in response to feeding with seaweed extracts. Proceedings of Sixth International Seaweed Symposium, Tokyo, (Blunden G (Ed.), 682-686.

Chapman GJ. 1970. Seaweed and their uses. Methuen and Company Limited, London.

Dhargalkar K and Untawale AG. 1983. Some observations of the effect of Seaweed Liquid Fertilizer on higher plants. Indian Journal of Marine Sciences 12:210-214.

Gandhiyappan K and Perumal P. Growth promoting effect of seaweed liquid fertilizer (*Enteromorpha intestinalis*) on the sesame crop plant. Seaweed Research and Utilization 23:23-25.

Kannan L and Tamilselvan C. 1990. Effect of seaweed manures on *Vigna radiatus*. In: Prof. MOP Iyenger Centenary Celebration India, Perspectives in Phycology (Rajarao VN., Ed.) 427-430.

Lowry OH, Rosenbrough NJ, Farr AL and Randall RJ. 1951. Protein measurement with the folin phenol reagent. Journal of Biological Chemistry 193:265-275.



Mackinney G. 1941. Absorption of light by chlorophyll solutions. Journal of Biological Chemistry 140:315-322.

Mohan VR, Venkataraman V, Murugeswari R and Muthuswami S. 1994. Effect of crude and commercial seaweed extracts on seed germination and seedling growth in *Cajanus cajan* L. Phykos 33:47-51.

Moore S and Stein WH. 1948. Photometric method for use in the chromatography amino acids. Journal of Biological Chemistry 176:367-388.

Nelson N. 1944. A photometric adoption of Somogyis method for the determination of reducing sugar. Journal of Biological Chemistry 31:426-428.

Rajkumar IS and Subramanian SK. 1999. Effect of fresh extracts and seaweed liquid fertilizers on some cereals and millets. Seaweed Research and Utilization 21:91-94.

Ramarao K. 1991. Effect of seaweed extract on *Zizyphus mauratiana* Lamk. Journal of Indian Botanical Society 71:19-21.

Tamilselvan C and Kannan L. 1994. Studies on the utilization of seaweeds as fertilizer for black gram. Indian Journal of Agricultural Research 28:121-126.

Taylor IEP and Wikinson AJ. 1977. The occurrence of gibberellins and gibberellins like substances in algae. Journal of Phycology 16:37-42.

Thivy F. 1961. Seaweed manure for perfect soil and soiling welds. Salt Research Industry 1:1-4.

Venkataraman KV and Mohan VR. 1997. Effect of seaweed liquid fertilizer on black gram. Phykos 36:43-47.

Venkataraman KV, Mohan VR, Murugeswari R and Muthuswamy M. 1993. Effects of crude and commercial seaweed extracts on seed germination and seedling growth in green gram and black gram. Seaweed Research and Utilization 16:23-27.

Wallwn KJO. Treasure form the sea. Organic Gardening 2:52-53.

